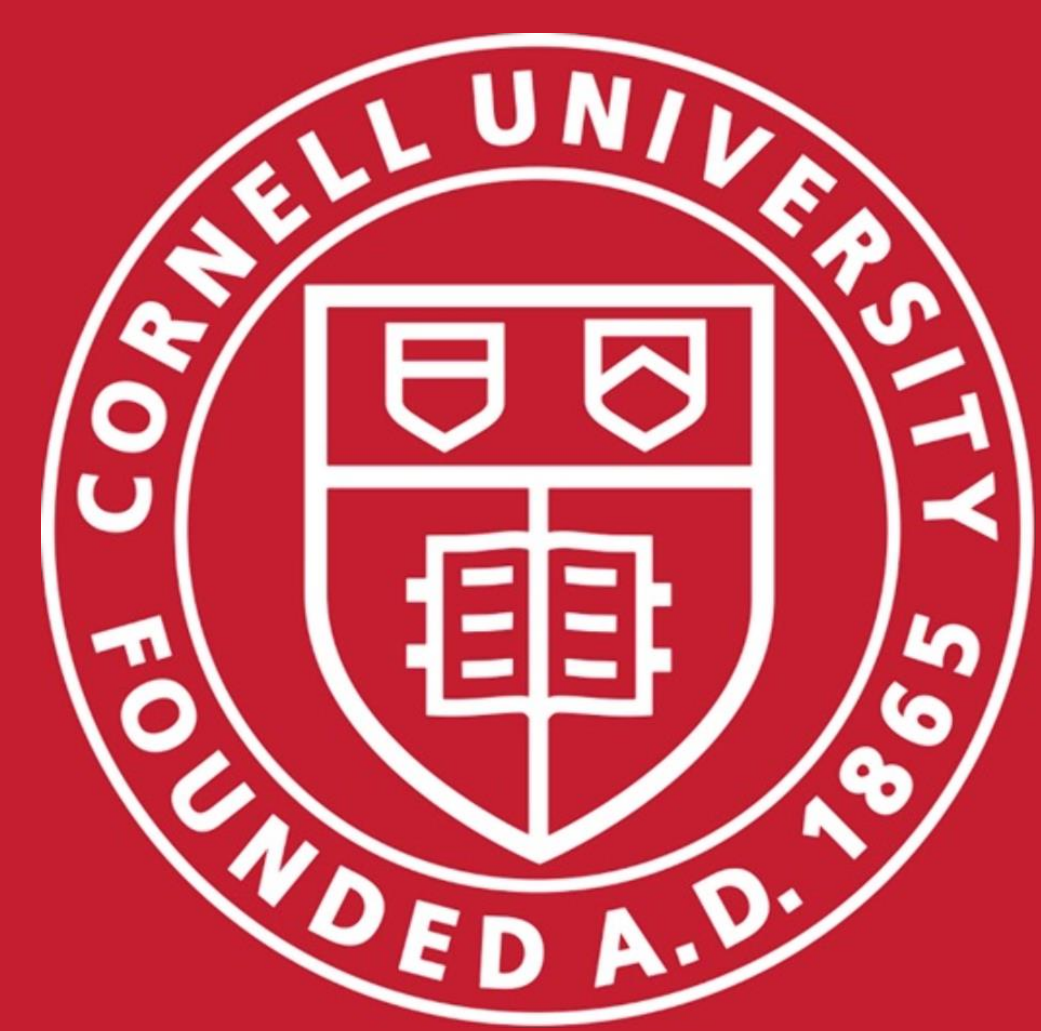




# Volatile Profiling Reveals Conserved VOC Responses Associated with Early *Pseudomonas syringae* pv. *aptata* Colonization of Beet Seeds

Muhammad Ali Mumtaz<sup>1</sup>, Lucas Foster Seybert<sup>2</sup>, Sean Patrick Murphy<sup>3</sup>, Rebecca Anne King<sup>1</sup>, Sarah Jane Pethybridge<sup>3</sup>, Christophe Duplais<sup>2</sup>, Collins Bugingo<sup>1</sup>.

<sup>1</sup>School of Integrative Plant Science, Horticulture Section, Cornell AgriTech, Cornell University, Geneva, NY 14456.

<sup>2</sup>Department of Entomology, Cornell AgriTech, Cornell University, Geneva, NY 14456.

<sup>3</sup>School of Integrative Plant Science, Plant Pathology and Plant-Microbe Biology Section, Cornell AgriTech, Cornell University, Geneva, NY 14456.

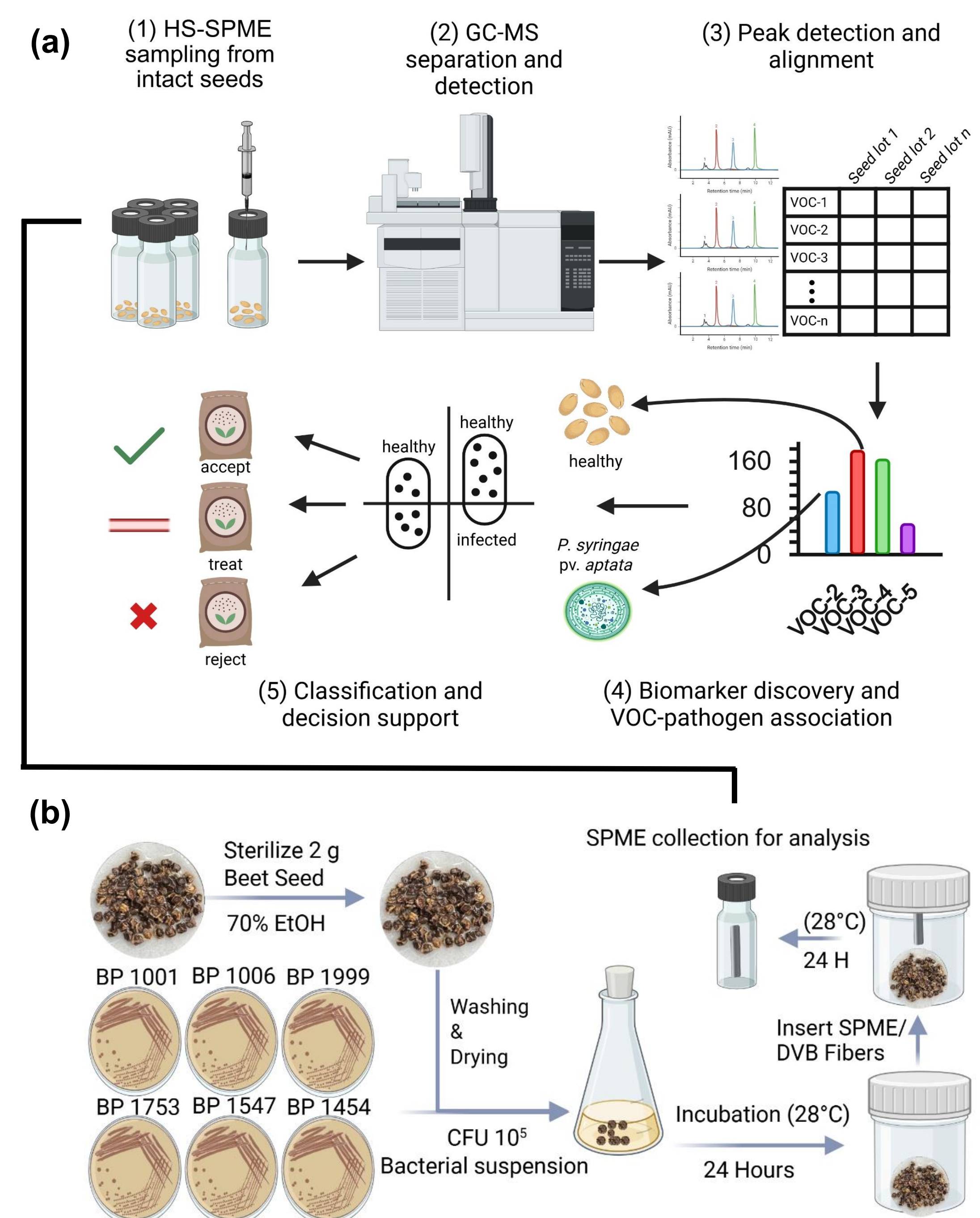
## Introduction

- Pseudomonas syringae* pv. *aptata* (Psa), the causal agent of bacterial leaf spot, is a genetically diverse, seedborne pathogen. Its ability to colonize seeds asymptotically facilitates its undetected introduction into crop ecosystems.
- Genetically distinct bacterial pathogens can produce distinct volatile fingerprints that reflect differences in their metabolism and physiological state (García-Barceló et al., 2026; Fitzgerald et al., 2020).
- Beyond serving as metabolic fingerprints, microbial VOCs mediate biological interactions, influence virulence, and enable non-destructive detection of seedborne pathogens in asymptomatic seeds (Wang et al., 2023; Gaudioso et al., 2025)."

## Objectives

- Characterize conserved and strain-specific volatile signatures associated with early Psa colonization of beet seed.
- Identify candidate VOC biomarkers for early detection and bioactive compounds with potential biocontrol activity.

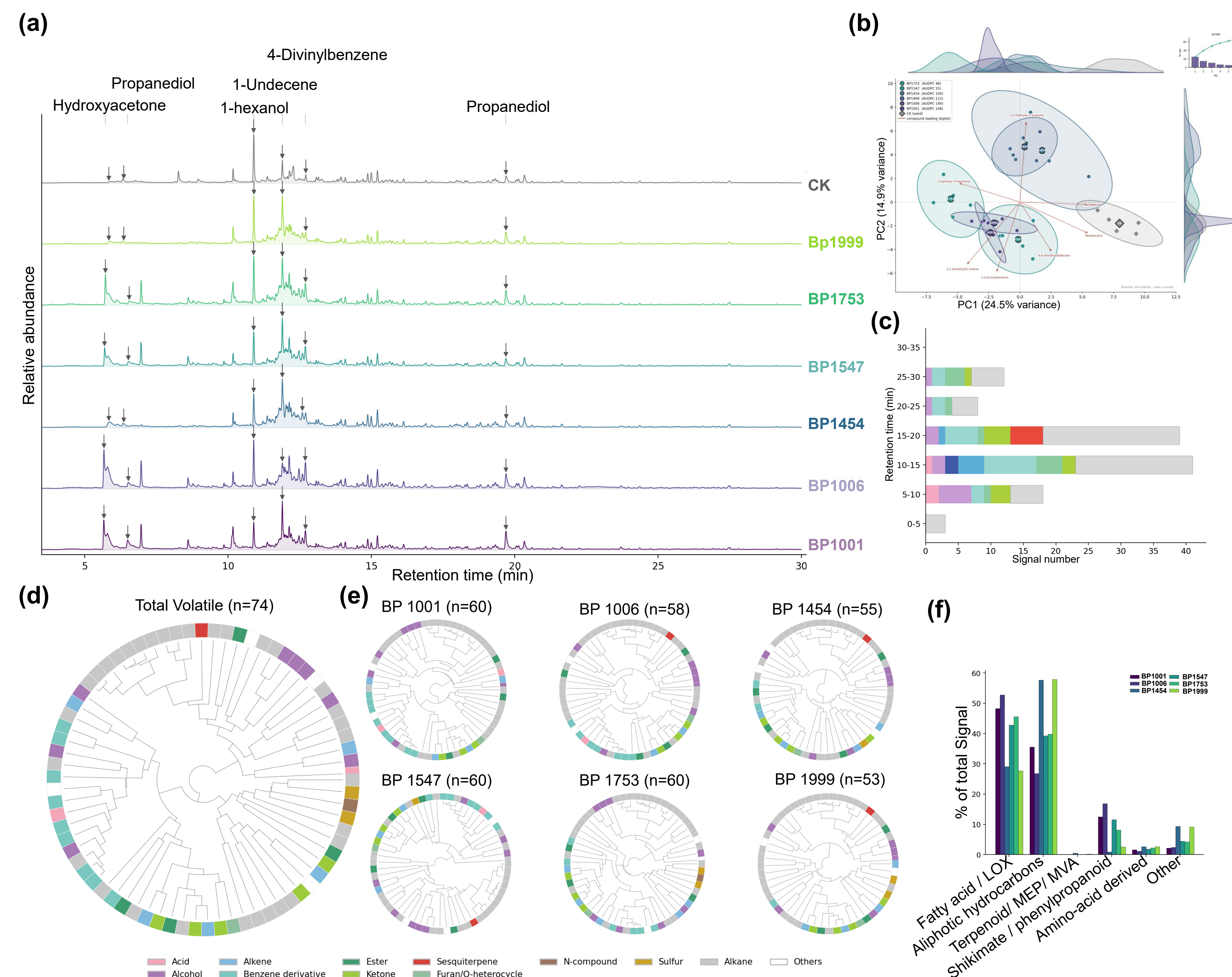
## Experimental Workflow and Methods



**Figure 1 Experimental Workflow and Methods.** (a) HS-SPME-GC-MS workflow for VOC collection, detection, and biomarker discovery in intact beet seeds. Identified VOC signatures can distinguish healthy from Psa-colonized seeds and support accept, treat, or reject decisions. (b). Experimental workflow for Psa inoculation and VOC collection. Surface-sterilized beet seeds were inoculated with six Psa strains, incubated at 28°C for 24 h, and sampled using SPME/DVB fibers for GC-MS analysis.

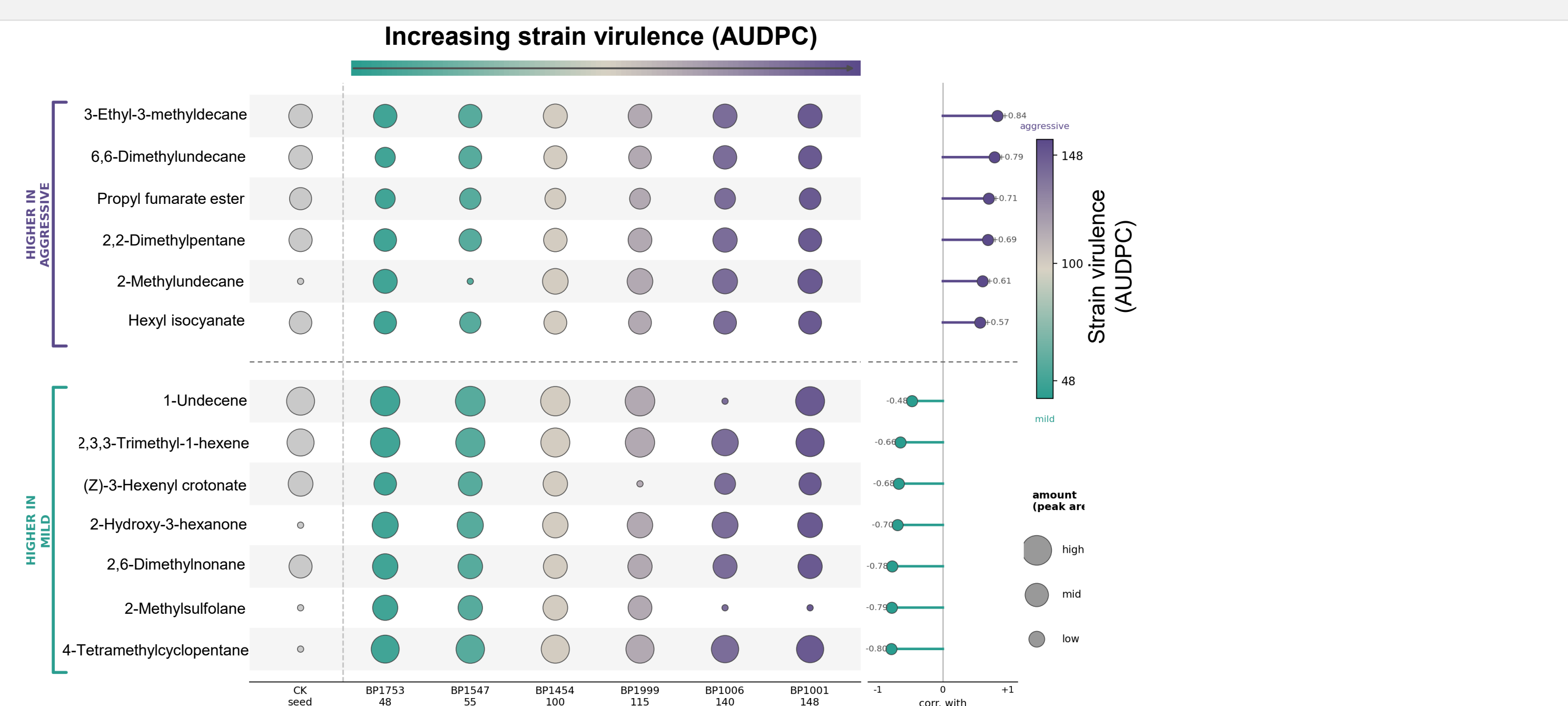
## Results

### Psa Strains Produce Distinct Volatilomic Fingerprints



**Figure 2. Strain-dependent VOC fingerprints of Psa-colonized beet seed** (a) Stacked GC-MS chromatograms comparing CK and six Psa strains (b) PCA biplot showing treatment-specific clustering and marginal distributions. (c) VOC abundance and chemical-class distribution across retention-time intervals. (d-e) Classification of all 74 VOCs and strain-specific subsets. (f) Relative contributions of putative biosynthetic pathways to total signal. Overall, Psa altered VOC identity, abundance, and biochemical origin in a strain-dependent manner. CK, uninoculated seed control.

### Virulence-Ranked VOC Signatures Associated with Psa



**Figure 3. Virulence-Ranked VOC Signatures Associated with Psa.** AUDPC values from Osabutey et al. (2025) were used for correlation analysis with VOC abundance. VOC abundance shifted with strain virulence: six compounds increased with AUDPC ( $r = 0.57-0.84$ ), whereas seven were enriched in milder strains ( $r = -0.48$  to  $-0.80$ ). These opposing patterns reveal distinct volatile signatures associated with aggressive and mild Psa strains.

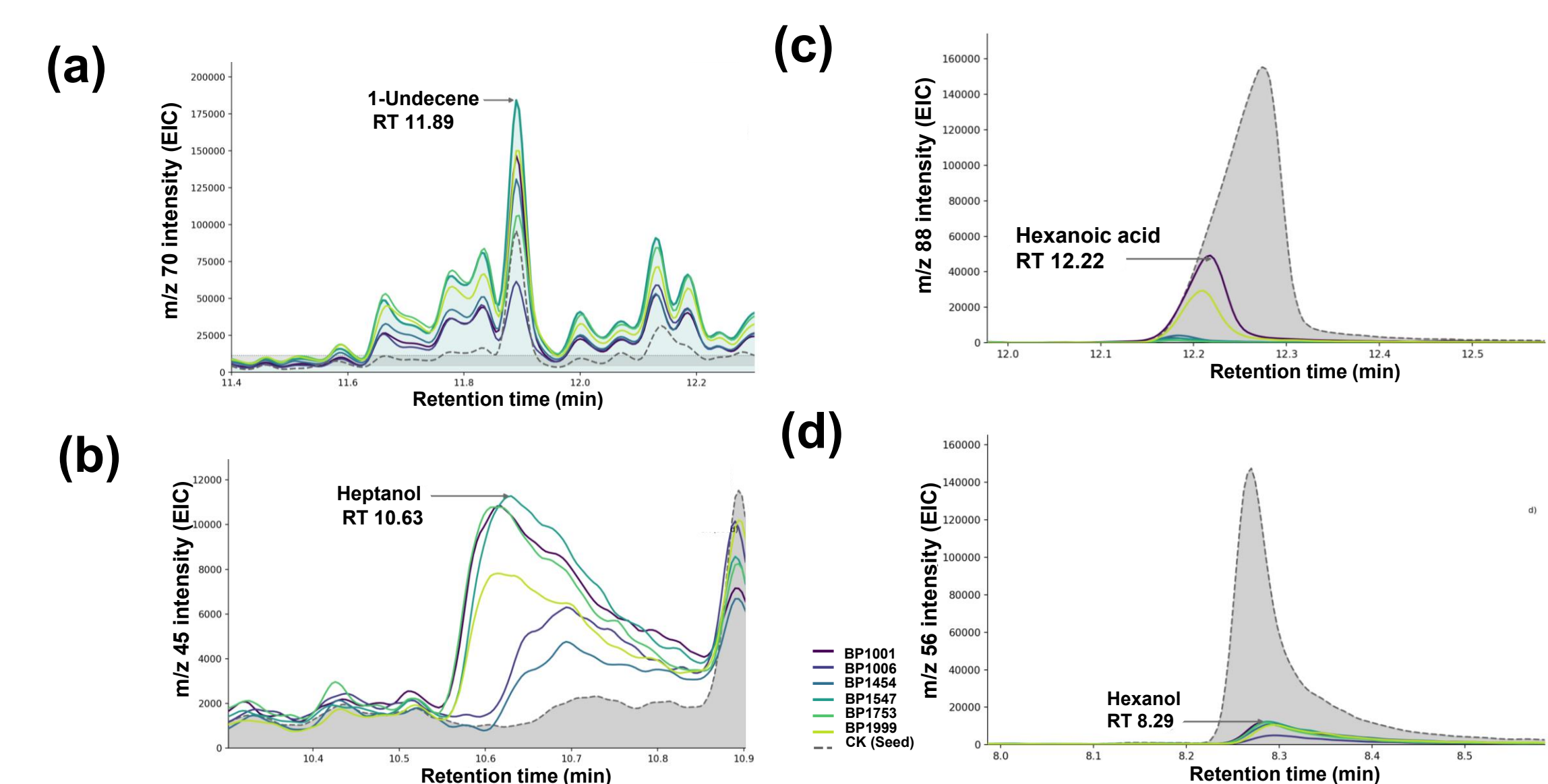
## Discussion

- Psa inoculation altered the Pablo seed volatilome. PCA separated the seed control from inoculated samples, while strain-specific clustering showed variation among Psa strains.
- The 74 detected VOCs covered diverse chemical classes, with fatty acid/LOX and aliphatic hydrocarbon signals contributing the largest proportions.
- 1-Undecene and 3-methyl-2-heptanol increased after inoculation, although their abundance varied among strains. In contrast, 1-hexanol and 2-ethylhexanoic acid decreased, indicating bacterial consumption, transformation, or altered seed emissions.

## Extracted-ion signal

### Candidate Biomarker

### Candidate Biocontrol



**Figure 4. Extracted-ion chromatograms of candidate VOCs.** (a-b) 1-Undecene and heptanol were enriched in Psa-inoculated seeds, supporting their use as candidate biomarkers. (c-d) Hexanoic acid and hexanol were depleted relative to CK and selected for further biocontrol evaluation.

## Key findings

- Psa colonization produced a clear, strain-dependent shift in the beet seed volatilome, revealing a multi-VOC signature separating inoculated seeds from uninoculated controls.
- A multi-VOC signature, including 1-undecene and 3-methyl-2-heptanol, showed potential for early detection of Psa-colonized seed.
- These findings support headspace volatilomics as a promising tool for identifying Psa-colonized seed and prioritizing compounds for biocontrol testing.

## Future Directions

- Validate candidate VOC biomarkers in a Psa-resistant beet genotype to assess their robustness across host genetic backgrounds.
- Evaluate identified VOCs for biocontrol activity against Psa and their ability to reduce seed colonization and disease development.

## Citations

- García-Barceló et al. (2026). doi.org/10.1038/s41598-026-58199-2
- Wang et al. (2023). doi.org/10.1038/s41396-023-01356-6
- Gaudioso et al. (2025). doi.org/10.3390/agronomy15020497
- Osabutey et al (2025). APS Plant Health.

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